



# Deterioration of apatite orientation in the cholecystinin B receptor gene (Cckbr)-deficient mouse femurs

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## Abstract

**Introduction** The discrepancy between bone mineral density (BMD), the gold standard for bone assessment, and bone strength is a constraint in diagnosing bone function and determining treatment strategies for several bone diseases. Gastric hypochlorhydria induced by clinically used proton pump inhibitor (PPI) therapy indicates a discordance between changes in BMD and bone strength. Here, we used Cckbr-deficient mice with gastric hypochlorhydria to examine the effect of gastric hypochlorhydria on bone mass, BMD, and preferential orientation of the apatite crystallites, which is a strong indicator of bone strength.

**Materials and methods** Cckbr-deficient mice were created, and their femurs were analyzed for BMD and preferential orientation of the apatite *c*-axis along the femoral long axis.

**Results** Cckbr-deficient mouse femurs displayed a slight osteoporotic bone loss at 18 weeks of age; however, BMD was comparable to that of wild-type mice. In contrast, apatite orientation in the femur mid-shaft significantly decreased from 9 to 18 weeks. To the best of our knowledge, this is the first report demonstrating the deterioration of apatite orientation in the bones of Cckbr-deficient mice.

**Conclusion** Lesions in Cckbr-deficient mice occurred earlier in apatite orientation than in bone mass. Hence, bone apatite orientation may be a promising method for detecting hypochlorhydria-induced osteoporosis caused by PPI treatment and warrants urgent clinical applications.

**Keywords** Cckbr-deficient mice · Bone quality · Apatite orientation · Hypochlorhydria · Bone evaluation

## Introduction

The dissociation between bone mineral density (BMD) and bone strength has been reported in both human and animal bones subjected to bone disorders, including fracture healing [1, 2], osteoporosis [3, 4], osteopetrosis [5], rheumatoid bone [6, 7], and metastasis [8]. These studies demonstrated

that bone quantity measurement alone is not sufficient to determine bone strength. To clarify the dissociation between bone mineral quantity and bone strength, extracellular matrix (ECM) quality should be considered [9]. Previously, we reported that the preferential orientation of the ECM, which is mainly composed of organic collagen fibers and inorganic apatite crystallites, is an important determinant of bone material integrity [10, 11]. Collagen and apatite exhibit anisotropic mechanical properties; the collagen fiber direction and apatite crystallographic *c*-axis display the highest strength. In the normal mineralization of bone, the crystallographic *c*-axis of apatite is aligned approximately parallel to the collagen fiber direction. This results in an oriented nanocomposite in which the strong directions of the two materials are aligned [12, 13]. Highly oriented apatite *c*-axes and collagen fibers mainly contribute to bone stiffening [11] and toughening [14], respectively. Therefore, evaluation of

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the preferential orientation of the bone ECM is important when discussing the fragility of pathological bones.

More than 20% of osteoporotic fracture patients were treated with proton pump inhibitors (PPI) to reduce the risk of the gastrointestinal side effects of bisphosphonates [15], which are the commonest medicines for osteoporosis. However, the negative effects of hypochlorhydria on bone properties were investigated using *Cckbr*-deficient mice [16]. *Cckbr*-deficient mice exhibit gastric hypochlorhydria and calcium malabsorption. Calcium deficiency leads to secondary hyperparathyroidism and excessive bone resorption, resulting in an osteoporotic phenotype. *Cckbr*-deficient mice demonstrate osteoporotic changes, characterized by an increase in osteoclasts on the bone surface, loss of bone volume and cortical bone thickness, elevated concentrations of bone-specific collagen degradation products, and decreased power endurance [16]. *Cckbr*-deficient mice showed secondary hyperparathyroidism at as early as 2 weeks of age, a marked decrease in BV/TV of the vertebral body at 12 weeks, and severe cortical porosity even in the cortical bone of tibia and femur at 52 weeks of age [16].

Osteoporosis has been reported to cause not only a decrease in BMD but also changes in ECM orientation that also significantly affects bone strength [17]. We hypothesized that ECM orientation is affected by *Cckbr*-deficiency-induced osteoporotic conditions. Therefore, the purpose of this novel study was to evaluate the preferential orientation of ECM as a bone quality parameter in *Cckbr*-deficient and wild-type mice. Since cases of ECM orientation being more acutely altered than BMD was in response to physiological changes have been reported [18], 9- and 18-week-old mice, before and after 12 weeks of age, when BV/TV was reduced, were used in this study. If a steeper change in ECM orientation than that in BMD is observed, it would be clinically significant from the perspective of the early detection of bone abnormalities. Additionally, gastric hypochlorhydria is caused by long-term proton pump inhibitor (PPI) therapy and is associated with an increased risk of osteoporosis [19]. Therefore, the analysis of these mice has important clinical implications.

## Materials and methods

### Animals

J1 embryonic stem (ES) cells were electroporated with a linearized targeting vector and selected with geneticin (G418) on embryonal fibroblast feeder cells. In total, 1033 G418-resistant clones were screened by Southern blot analysis using 5' external and 3' internal probes. Six clones showed evidence of homologous recombination of the disrupted *Cckbr* gene. Four ES clones were microinjected into

the blastocysts of C57BL/6 J female mice. Two independent ES clones generated germline chimeras. Chimeras were bred with C57BL/6 J and 129sv mice to generate heterozygous mutant F1 mice [20]. We purchased these mice from the RIKEN BioResource Center. All the mice were housed in a colony room controlled for temperature and humidity with a 12:12-h light/dark cycle (lights on at 07:00 h) with food, 5001-Laboratory Rodent Diet (Land O'Lakes, MN, US), and water available ad libitum. Twenty mice (10 *Cckbr*-deficient and 10 wild-type) were used for the analyses. At 9 and 18 weeks of age ( $N=5$ ), the animals were sacrificed using an overdose of sodium pentobarbital, and the right femur was harvested and immersed in a 75% ethanol solution to prevent denaturation of the organic constituents. The fourth lumbar vertebra was similarly sampled for the comparison of BMD changes with the femur.

All experiments were conducted according to protocols approved by the Animal Care and Use Committee of Hamamatsu University School of Medicine (approval number: 2019002).

### Radiography

Soft X-ray photographs (XIE; Chubu Medical, Mie, Japan) were acquired after the femur was removed using 30 kV and 30  $\mu$ A radiation.

### Evaluation of volumetric BMD and cross-sectional area

Volumetric BMD and cross-sectional area were measured using a peripheral quantitative computed tomography (pQCT) (XCT Research SA + system; Stratec Medizintechnik GmbH, Birkenfeld, Germany) with a resolution of  $70 \times 70 \times 260 \mu\text{m}$ . This analysis was performed at regular intervals of 1/10th the bone length along the bone axis. The bone tissue was arbitrated to be above the threshold value of  $690 \text{ mg/cm}^3$ . In addition, the volumetric BMD of the transverse section at the height center of the fourth lumbar vertebra was measured under the same conditions. The threshold for bone tissue was set at  $395 \text{ mg/cm}^3$ , which is defined as the threshold for cancellous bone.

### Analysis of apatite *c*-axis orientation (bone quality)

Because the apatite *c*-axis is oriented almost parallel to the collagen fiber direction, the apatite *c*-axis orientation mirrors the collagen fiber orientation [12, 13]. The degree of apatite *c*-axis orientation was analyzed using a microbeam X-ray diffractometer ( $\mu$ XRD) (R-Axis BQ; Rigaku, Tokyo, Japan) equipped with a transmission-type optical system and an imaging plate (storage phosphors) (Fuji Film, Tokyo, Japan) placed behind the specimen. Mo-K $\alpha$

radiation with a wavelength of 0.07107 nm was generated at a tube voltage of 50 kV and a tube current of 90 mA. The distance between the detector and the X-ray focus of the specimen was 127.4 mm and the pixel size of the imaging plate was  $100\ \mu\text{m} \times 100\ \mu\text{m}$ . Subsequently, the incident beam was radiated vertically to the long axis of the bone to detect diffraction along the bone axis. It was focused on a beam spot of  $300\ \mu\text{m}$  in diameter by a double-pinhole metal collimator and radiated in the anteroposterior axis from the anterior surface at the center of the bone width (Fig. 1a). The diffraction data were collected for 300 s. The analysis was performed at positions where the  $\mu\text{XRD}$  analysis was implemented. From the obtained diffraction intensity pattern (Debye ring) (Fig. 1b), two representative diffraction peaks of apatite (002) and (310) were used for apatite *c*-axis orientation analysis [21]. In long bones, the apatite *c*-axis is preferentially oriented along the longitudinal axis of the bone aligned with the collagen matrix [13]. Therefore, in this study, diffraction information along the long axis of the femur was analyzed. The upper and lower parts of the Debye ring correspond to the long axis of the femur (Fig. 1b). The diffraction intensities were azimuthally integrated into the range of 100 pixels to obtain an X-ray diffraction profile.

The degree of the preferential orientation of the apatite *c*-axis was determined as the relative intensity ratio of the (002) diffraction peak to the (310) peak in the X-ray profile. This has been reported as a suitable index for evaluating apatite orientation [10, 11]. Subsequently, the intensity ratios calculated for the upper and lower parts of the Debye ring were averaged. Randomly oriented hydroxyapatite

(NIST #2910: calcium hydroxyapatite) powder had an intensity ratio of 0.8; therefore, the detected values  $> 0.8$  indicated the presence of anisotropic apatite *c*-axis orientation in the analyzed direction.

## Statistical analyses

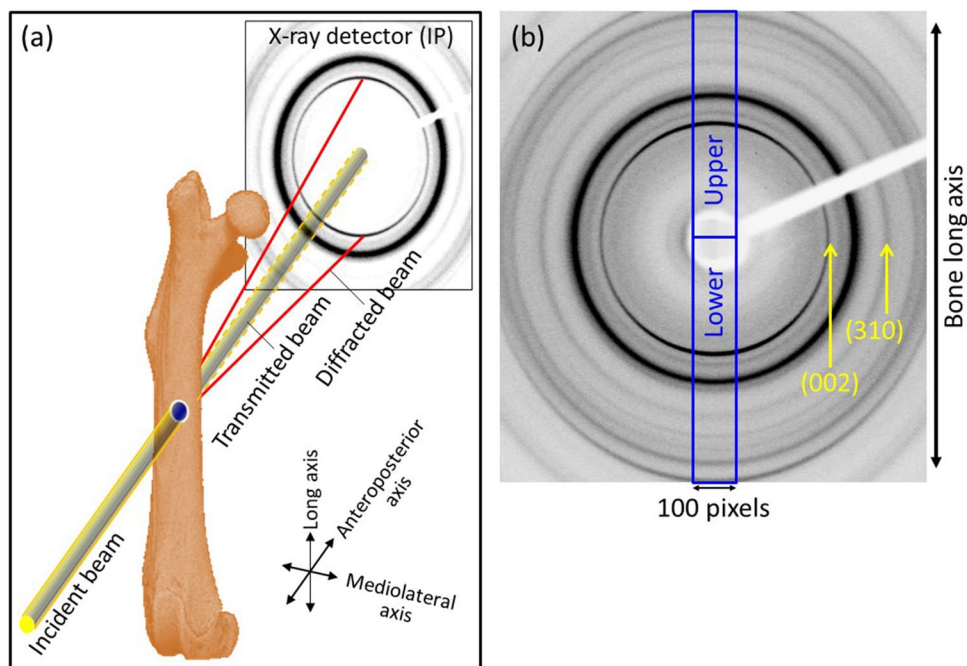
Quantitative results are expressed as mean  $\pm$  the standard deviation (SD). A two-tailed *t* test was used to compare data between Cckbr-deficient and wild-type mice.  $P < 0.05$  was considered significant. SPSS version 25 software (SPSS Japan Inc., Tokyo, Japan) for Microsoft Windows was used for all statistical analyses.

## Results

### Shape of the femur

Figure 2 reveals the soft X-ray photographs of Cckbr-deficient and wild-type mice at 9 and 18 weeks of age. At 9 weeks, there were no obvious differences in the shape of the femoral bone between Cckbr-deficient and wild-type mice. In contrast, at 18 weeks of age, the femoral cortex appeared thin in Cckbr-deficient mice (Fig. 2). Cckbr-deficient mice had a significantly thinner cross-sectional femoral bone area at 18 weeks of age, whereas no significant differences were detected at 9 weeks of age (Fig. 3).

**Fig. 1**  $\mu\text{XRD}$  analysis is performed in this study. **a** Schematic drawing of an optical system with the bone specimen and **b** a typical obtained  $\mu\text{XRD}$  pattern (Debye ring)



## BMD

Cortical BMD minimally differed between Cckbr-deficient and wild-type female mouse femurs at 9 and 18 weeks of age, as shown in Fig. 4. On the other hand, vertebral BMD was significantly lower in Cckbr-deficient mice at 18 weeks of age (Fig. 5).

## Apatite *c*-axis orientation along the bone long axis

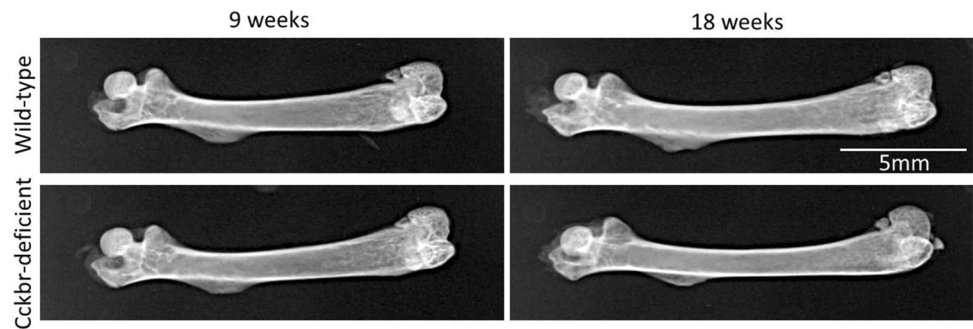
Figure 6a depicts the degree of the apatite *c*-axis orientation, analyzed as the intensity ratio of (002)/(310) along the long bone axis. In the femurs of wild-type mice, the

degree of apatite orientation peaked at the femoral mid-shaft and decreased toward the metaphysis. In contrast, Cckbr-deficient mouse femurs demonstrated a significantly lower degree of apatite *c*-axis orientation than that demonstrated by wild-type mouse femurs around the mid-shaft.

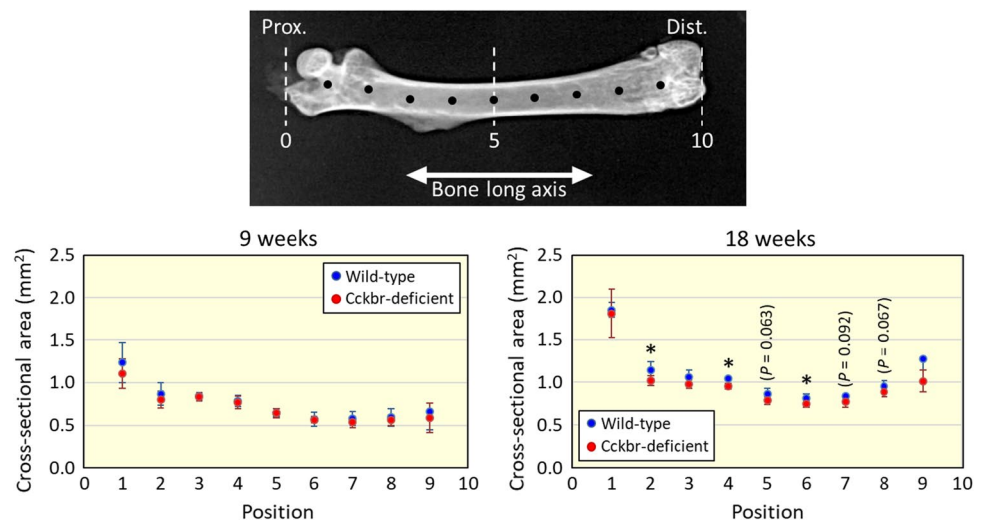
## Discussion

This study measured and compared the density and preferential orientation of apatite at the bone tissue level in the femurs of Cckbr-deficient and wild-type mice. These measurements were performed on the cortical bone at the

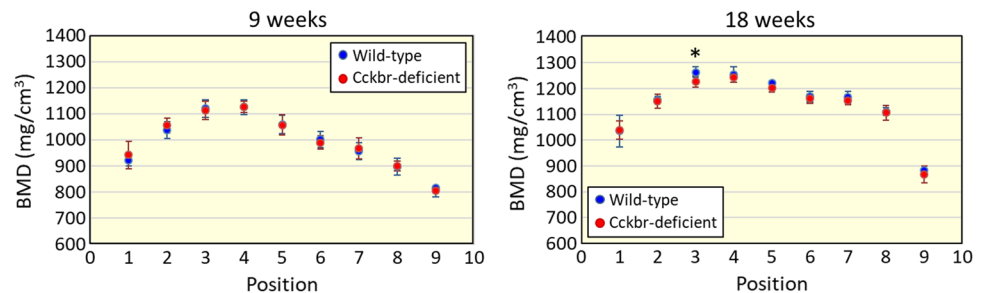
**Fig. 2** Soft X-ray photographs of Cckbr-deficient and wild-type mouse femurs at 9 and 18 weeks of age

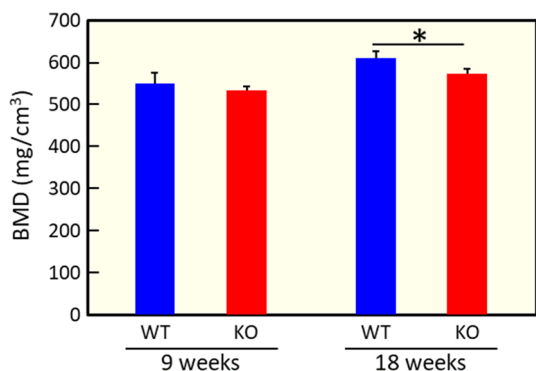


**Fig. 3** Distribution of cross-sectional bone area of the femur along the bone long axis for Cckbr-deficient and wild-type mice. Analyzed points are indicated in soft X-ray photographs. \* $P < 0.05$



**Fig. 4** Distribution of BMD of the femur along the bone long axis. \* $P < 0.05$





**Fig. 5** BMD of the fourth lumbar vertebra in the central transverse section. \**P* < 0.05

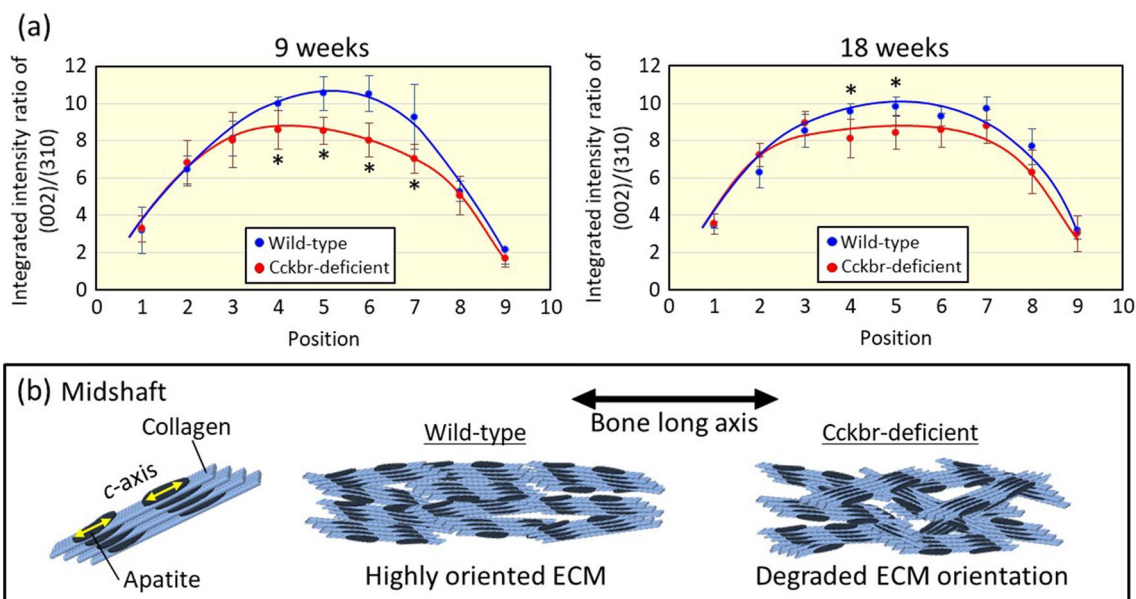
mid-shaft of the femur. Our findings indicated a deterioration in apatite orientation in the femurs of *Cckbr*-deficient mice, without significant differences in volumetric BMD.

The volumetric BMD of the femoral diaphysis, which is composed almost entirely of cortical bone, is extremely sluggish in its response to metabolic disease compared to that associated with the spinal BMD [22]. In fact, vertebral BMD showed a decreasing trend at 9 weeks and was significantly lower at 18 weeks of age. Since rodent cortical bone lacks osteons and does not undergo remodeling, bone formation and resorption occur primarily at the bone surface. Therefore, in this study, the effect of osteoporotic change appeared as a slight reduction in cross-sectional area and

not in volumetric BMD. Invariance of BMD in the long bone cortex due to osteoporosis has also been reported in an OVX model [18]. With further aging of *Cckbr*-deficient mice (e.g., at 52 weeks of age), volumetric BMD should decrease as severe pore formation inside cortical bone occurs [16].

Cholecystokinin (CCK) is a gut–brain peptide that exerts various physiological effects in the gastrointestinal tract and nervous system through cell-surface CCK receptors [23, 24]. CCK receptors have been divided into two subtypes: the CCK-A and CCK-B receptors (CCKBR), both belonging to the class of G protein-coupled receptors characterized by seven transmembrane domains [24].

One role of CCKBR [24–26] includes the stimulation of gastric acid secretion via parietal cells by binding the endogenous peptide hormone gastrin [27, 28]. This has been experimentally confirmed in *Cckbr*-deficient mice, which mimic gastric hypochlorhydria [16]. *Cckbr*-deficient mice display gastric hypochlorhydria and calcium malabsorption. Further, calcium deficiency leads to secondary hyperparathyroidism and excessive bone resorption, resulting in an osteoporotic bone phenotype [16]. However, femoral BMD did not differ between *Cckbr*-deficient and wild-type mice in this study. In contrast, the apatite orientation significantly deteriorated in *Cckbr*-deficient mice. A previous study reported impaired apatite orientation under hyperparathyroidism in rats with chronic kidney injury [29], and it is possible that hyperparathyroidism itself may have influenced the impaired orientation. However, since uremia also occurred in this model, it should not be concluded that the



**Fig. 6** Comparison of apatite orientation (bone quality parameter). **a** Variation in the preferential orientation of apatite *c*-axis along bone long axis quantitatively analyzed by the intensity ratio of (002) and

(310) diffraction peaks and **b** schematic representation of ECM micro-organization for *Cckbr*-deficient and wild-type mouse femur. \**P* < 0.05

decreased orientation was caused by hyperparathyroidism. Therefore, factors other than hyperparathyroidism that may have contributed to the impaired apatite orientation need to be discussed.

Being mechanosensors, osteocytes are affected by  $\text{Ca}^{2+}$  signals [30]. Considering that osteocyte morphology and alignment are necessary for the formation of ECM orientation [31], hypocalcemia induced by hypochlorhydria impairs the formation of a preferential ECM orientation. In contrast, a negative correlation has been reported between bone matrix anisotropy and regional periosteal mineral apposition rates [32]. Reports indicated the difference in recovery between BMD and preferential apatite *c*-axis [11], suggesting a discrepancy between the ECM orientation and BMD in this study.

Although BMD measurement did not detect the osteoporotic change due to hypochlorhydria, the assessment of bone quality using ECM orientation did. PPI, which induces hypochlorhydria, is clinically used for gastroesophageal reflux disease, Barrett's esophagus, and non-steroidal anti-inflammatory drug-related bleeding prophylaxis [19]. Therefore, the investigation of PPI-induced osteoporosis is crucial.

In contrast, CCKBR induces pain sensitivity [33, 34]. CCKBR antagonists restored the effectiveness of morphine [35] and attenuated the symptoms of mechanical allodynia in a neuropathic pain model [36]. The upregulation of the opioid system [37] and elevated thresholds of nociceptive stimuli [38] have been observed in *Cckbr*-deficient mice. Moreover, the deletion of *Cckbr* reduces mechanical sensitivity [39]. Under unloading conditions, the degree of ECM orientation along the loaded direction significantly degraded [21]. Therefore, mechanical sensitivity may affect the formation of preferential ECM orientations. We considered that the reduction of a specific loading direction of the limbs due to the loss of mechanical sensitivity induced the deterioration of ECM orientation. In *Cckbr*-deficient mice, hypochlorhydria and deteriorated apatite orientation, which is possibly induced by mechanical hyposensitivity, occur simultaneously, and further investigation is needed to elucidate their direct relationship. Clinically, mechanical sensitivity disorders and specific neuropathies, are induced by diabetes mellitus [40] and spinal disorders [41]. Thus, it is important to examine the effect of mechanical sensitivity on the formation of ECM orientation.

In this study, the deteriorated apatite orientation in *Cckbr*-deficient mouse femurs partially recovered between 9 and 18 weeks of age. In contrast, the cross-sectional bone area was significantly decreased in 18-week-old but not 9-week-old *Cckbr*-deficient mice. This decrease in the cross-sectional bone area may compensate for the deterioration of apatite orientation in *Cckbr*-deficient mice.

Bone changes due to *Cckbr*-deficiency-induced hypochlorhydria scarcely appeared in bone density throughout the

experimental period, appeared in the cross-sectional area (bone mass) at 18 weeks of age, and were detected in the apatite orientation around the mid-shaft as early as 9 weeks of age. This finding provides insight into the early detection of unfavorable bone changes under PPI treatment, which is widely used to induce hypochlorhydria. However, it is necessary to clarify the differences between congenital genetic defects and acquired drug treatments. The method used in this study to analyze the apatite orientation is invasive because bone extraction was required. Recently, a non-invasive method using ultrasound imaging was developed [42], which may be useful for the clinical diagnosis of hypochlorhydria.

In conclusion of this study, we analyzed cross-sectional bone area, BMD, and apatite orientation (a bone quality parameter) in the femoral cortex of *Cckbr*-deficient and wild-type mice. Deterioration of the apatite preferential orientation in the femur mid-shaft of *Cckbr*-deficient mice was demonstrated for the first time, despite unchanged BMD. Our results suggest that hypochlorhydria and impaired mechanical sensitivity of *Cckbr*-deficient mice are at least partially attributable to the degradation of apatite orientation. Moreover, apatite orientation is beneficial in detecting hypochlorhydria induced by PPI treatment, whereas BMD is insensitive.

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**Author contributions** YM and TN conceptualized and supervised the study and reviewed and edited the manuscript. RO, TO, YY, TY, and AO collected the data. YM and TI wrote the manuscript. All the authors have read and approved the final version of the manuscript.

## Declarations

**Conflict of interest** None.

**Ethical approval** This study was approved by the Animal Care and Use Committee of Hamamatsu University School of Medicine (approval number: 2019002).

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